# Package 'growthrates'

June 15, 2025

```
Encoding UTF-8
Type Package
Title Estimate Growth Rates from Experimental Data
Version 0.8.5
Date 2025-06-14
LazyData yes
Maintainer Thomas Petzoldt < thomas.petzoldt@tu-dresden.de>
Description A collection of methods to determine growth rates from
      experimental data, in particular from batch experiments and
      plate reader trials.
Depends R (>= 3.2), lattice, deSolve
Imports stats, graphics, methods, parallel, utils, FME
Suggests knitr, rmarkdown, dplyr, ggplot2
License GPL (>= 2)
URL https://github.com/tpetzoldt/growthrates
VignetteBuilder knitr
RoxygenNote 7.3.2
Collate 'aaa growthmodel-class.R' 'aab growthmodel-constructor.R'
      'aac_classes.R' 'aad_set_generics.R' 'all_easylinear.R'
      'all growthmodels.R' 'all splines.R' 'antibiotic.R'
      'bactgrowth.R' 'cost.R' 'extcoef_logistic.R' 'extract.R'
      'fit_easylinear.R' 'fit_growthmodel.R' 'fit_spline.R'
      'grow_baranyi.R' 'grow_exponential.R' 'grow_genlogistic.R'
      'grow_gompertz.R' 'grow_gompertz2.R' 'grow_huang.R'
      'grow_logistic.R' 'grow_richards.R' 'grow_twostep.R'
      'growthrates-package.R' 'lm_window.R' 'methods.R'
      'multisplit.R' 'names.R' 'parse_formula.R'
      'parse_formula_nonlin.R' 'plot.R' 'predict.R' 'utilities.R'
NeedsCompilation yes
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```

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# Repository CRAN

**Date/Publication** 2025-06-15 17:30:02 UTC

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# Description

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A collection of methods to determine growth rates from experimental data, in particular from batch experiments and plate reader trials.

The package contains basically three methods:

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• fit a linear regression to a subset of data with the steepest log-linear increase (a method, similar to Hall et al., 2013),

- fit parametric nonlinear models to the complete data set, where the model functions can be given either in closed form or as numerically solved (system of) differential equation(s),
- use maximum of the 1st derivative of a smoothing spline with log-transformed y-values (similar to Kahm et al., 2010).

The package can fit data sets of single experiments or complete series containing multiple data sets. Included are functions for extracting estimates and for plotting. The package supports growth models given as numerically solved differential equations. Multi-core computation is used to speed up fitting of parametric models.

### Author(s)

Thomas Petzoldt

#### References

Hall, B. G., Acar, H. and Barlow, M. 2013. Growth Rates Made Easy. Mol. Biol. Evol. 31, 232-238, doi:10.1093/molbev/mst197

Kahm, M., Hasenbrink, G., Lichtenberg-Frate, H., Ludwig, J., Kschischo, M. 2010. grofit: Fitting Biological Growth Curves with R. Journal of Statistical Software, 33(7), 1-21, doi:10.18637/jss.v033.i07

Soetaert, K. and Petzoldt, T. 2010. Inverse Modelling, Sensitivity and Monte Carlo Analysis in R Using Package FME. Journal of Statistical Software, 33(3), 1-28, doi:10.18637/jss.v033.i03

Soetaert, K., Petzoldt, T. Setzer, R. W. 2010. Solving Differential Equations in R: Package deSolve. Journal of Statistical Software, 33(9), 1-25, doi:10.18637/jss.v033.i09

#### See Also

```
fit_easylinear, fit_spline, fit_growthmodel, all_easylinear, all_splines, all_growthmodels
```

```
data(bactgrowth)
splitted.data <- multisplit(bactgrowth, c("strain", "conc", "replicate"))
## get table from single experiment
dat <- splitted.data[["D:0:1"]]

fit1 <- fit_spline(dat$time, dat$value)
plot(fit1, log="y")
plot(fit1)

## derive start parameters from spline fit
p <- coef(fit1)

## subset of first 10 data
first10 <- dat[1:10, ]</pre>
```

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```
fit2 <- fit_growthmodel(grow_exponential, p=p, time=first10$time, y=first10$value)

## use parameters from spline fit and take K from the data maximum

p <- c(coef(fit1), K = max(dat$value))

fit3 <- fit_growthmodel(grow_logistic, p=p, time=dat$time, y=dat$value, transform="log")

plot(fit1)
lines(fit2, col="green")
lines(fit3, col="red")</pre>
```

all\_easylinear

Easy Growth Rates Fit to data Frame

# Description

Determine maximum growth rates from log-linear part of the growth curve for a series of experiments.

# Usage

```
all_easylinear(...)
## S3 method for class 'formula'
all_easylinear(formula, data, h = 5, quota = 0.95, subset = NULL, ...)
## S3 method for class 'data.frame'
all_easylinear(
   data,
   grouping,
   time = "time",
   y = "value",
   h = 5,
   quota = 0.95,
   ...
)
```

### **Arguments**

generic parameters, reserved for future extensions.

formula model formula specifying dependent, independent and grouping variables in the

form: dependent ~ independent | group1 + group2 + ....

data data frame of observational data.

h with of the window (number of data).

quota part of window fits considered for the overall linear fit (relative to max. growth

rate).

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subset	a specification of the rows to be used: defaults to all rows.
grouping	model formula or character vector of criteria defining subsets in the data frame.
time	character vectors with name independent variable.
у	character vector with name of dependent variable

### Value

object with parameters of all fits.

### References

```
Hall, BG., Acar, H, Nandipati, A and Barlow, M (2014) Growth Rates Made Easy. Mol. Biol. Evol. 31: 232-38, doi:10.1093/molbev/mst187
```

### See Also

```
Other fitting functions: all_growthmodels(), all_splines(), fit_easylinear(), fit_growthmodel(), fit_spline()
```

# **Examples**

```
library("growthrates")
L <- all_easylinear(value ~ time | strain + conc + replicate, data=bactgrowth)
summary(L)
coef(L)
rsquared(L)
results <- results(L)
library(lattice)
xyplot(mumax ~ conc|strain, data=results)</pre>
```

all\_growthmodels

Fit Nonlinear Growth Models to Data Frame

### **Description**

Determine maximum growth rates by nonlinear fits for a series of experiments.

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### Usage

```
all_growthmodels(...)
## S3 method for class 'formula'
all_growthmodels(
  formula,
  data,
 p,
  lower = -Inf,
  upper = Inf,
 which = names(p),
 FUN = NULL,
 method = "Marq",
  transform = c("none", "log"),
  . . . ,
 subset = NULL,
 ncores = detectCores(logical = FALSE)
)
## S3 method for class '`function`'
all_growthmodels(
  FUN,
 p,
  data,
  grouping = NULL,
  time = "time",
  y = "value",
  lower = -Inf,
  upper = Inf,
 which = names(p),
 method = "Marq",
  transform = c("none", "log"),
 ncores = detectCores(logical = FALSE)
)
```

### **Arguments**

	generic parameters, including parameters passed to the optimizer.
formula	model formula specifying dependent, independent and grouping variables in the form: dependent ~ independent   group1 + group2 +
data	data frame of observational data.
р	named vector of start parameters and initial values of the growth model.
lower	lower bound of the parameter vector.
upper	upper bound of the parameter vector.
which	vector of parameter names that are to be fitted.

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FUN function of growth model to be fitted.

method character vector specifying the optimization algorithm.

transform fit model to non-transformed or log-transformed data.

subset a specification of the rows to be used: defaults to all rows.

ncores number of CPU cores used for parallel computation. The number of real cores is

detected automatically by default, but fort debugging purposes it could be wise to set ncores = 1. Usage of logical (hyperthreading) cores does not speed up

computation.

grouping vector of grouping variables defining subsets in the data frame.

time character vector with name of independent variable.
y character vector with name of dependent variable.

### Value

object containing the parameters of all fits.

#### See Also

```
Other fitting functions: all_easylinear(), all_splines(), fit_easylinear(), fit_growthmodel(), fit_spline()
```

```
data(bactgrowth)
splitted.data <- multisplit(value ~ time | strain + conc + replicate,</pre>
                  data = bactgrowth)
## show which experiments are in splitted.data
names(splitted.data)
## get table from single experiment
dat <- splitted.data[["D:0:1"]]</pre>
fit0 <- fit_spline(dat$time, dat$value)</pre>
fit1 <- all_splines(value ~ time | strain + conc + replicate,</pre>
                  data = bactgrowth, spar = 0.5)
## these examples require some CPU power and may take a bit longer
## initial parameters
p <- c(coef(fit0), K = max(dat$value))</pre>
## avoid negative parameters
lower = c(y0 = 0, mumax = 0, K = 0)
## fit all models
fit2 <- all_growthmodels(value ~ time | strain + conc + replicate,</pre>
```

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```
data = bactgrowth, FUN=grow_logistic,
          p = p, lower = lower, ncores = 2)
results1 <- results(fit1)</pre>
results2 <- results(fit2)</pre>
plot(results1$mumax, results2$mumax, xlab="smooth splines", ylab="logistic")
## experimental: nonlinear model as part of the formula
fit3 <- all_growthmodels(</pre>
          value ~ grow_logistic(time, parms) | strain + conc + replicate,
          data = bactgrowth, p = p, lower = lower, ncores = 2)
## this allows also to fit to the 'global' data set or any subsets
fit4 <- all_growthmodels(</pre>
          value ~ grow_logistic(time, parms),
          data = bactgrowth, p = p, lower = lower, ncores = 1)
plot(fit4)
fit5 <- all_growthmodels(</pre>
          value ~ grow_logistic(time, parms) | strain + conc,
          data = bactgrowth, p = p, lower = lower, ncores = 2)
plot(fit5)
```

all\_splines

Fit Exponential Growth Model with Smoothing Spline

### **Description**

Determine maximum growth rates from log-linear part of the growth curve for a series of experiments by using smoothing splines.

# Usage

```
all_splines(...)
## S3 method for class 'formula'
all_splines(formula, data = NULL, optgrid = 50, subset = NULL, ...)
## S3 method for class 'data.frame'
all_splines(
   data,
   grouping = NULL,
   time = "time",
   y = "value",
   optgrid = 50,
   ...
)
```

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### Arguments

• • •	generic parameters, including parameters passed to smooth.spline, see details.
formula	model formula specifying dependent, independent and grouping variables in the form: dependent $\sim$ independent   group1 + group2 +
data	data frame of observational data.
optgrid	number of steps on the x-axis used for searching the maximum of the first derivative of the spline. The default should work in most cases, as long as the data are equally spaced. A smaller number may lead to non-detectable speed-up, but has the risk that the search is trapped in a local minimum.
subset	a specification of the rows to be used: defaults to all rows.
grouping	vector of grouping variables defining subsets in the data frame.
time	character vectors with name independent variable.

#### **Details**

У

The method was inspired by an algorithm of Kahm et al. (2010), with different settings and assumptions. In the moment, spline fitting is always done with log-transformed data, assuming exponential growth at the time point of the maximum of its first derivative.

character vector with name of dependent variable.

All the hard work is done by function smooth.spline from package stats, that is highly user configurable. Normally, smoothness is automatically determined via cross-validation. This works well in many cases, whereas manual adjustment is required otherwise, e.g. by setting spar to a fixed value [0,1] that also disables cross-validation. A typical case where cross validation does not work is, if time dependent measurements are taken as pseudoreplicates from the same experimental unit.

# Value

object with parameters of the fit.

### References

Kahm, M., Hasenbrink, G., Lichtenberg-Frate, H., Ludwig, J., Kschischo, M. 2010. grofit: Fitting Biological Growth Curves with R. Journal of Statistical Software, 33(7), 1-21, doi:10.18637/jss.v033.i07

### See Also

```
Other fitting functions: all_easylinear(), all_growthmodels(), fit_easylinear(), fit_growthmodel(), fit_spline()
```

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antibiotic

Plate Reader Data of Bacterial Growth

# **Description**

Example data set from growth experiments with Pseudomonas putida on a tetracycline concentration gradient.

### **Format**

Data frame with the following columns:

```
time time in hours.
```

variable sample code.

value bacteria concentration measured as optical density.

conc concentration of the antibiotics (Tetracycline).

repl Replicate.

### **Details**

The sample data set shows four out of six replicates of the original experiment.

#### **Source**

Claudia Seiler, TU Dresden, Institute of Hydrobiology.

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### **Examples**

```
## plot data and determine growth rates
data(antibiotic)

dat <- subset(antibiotic, conc==0.078 & repl=="R4")
parms <- c(y0=0.01, mumax=0.2, K=0.5)
fit <- fit_growthmodel(grow_logistic, parms, dat$time, dat$value)
plot(fit); plot(fit, log="y")</pre>
```

bactgrowth

Plate Reader Data of Bacterial Growth

### **Description**

Example data set from growth experiments with different concentrations of antibiotics.

#### **Format**

Data frame with the following columns:

strain identifier of the bacterial strain, D=donor, R=recipient, T=transconjugant.

replicate replicate of the trial.

conc concentration of the antibiotics (Tetracycline).

time time in hours.

value bacteria concentration measured as optical density.

### **Details**

This rather 'difficult' data set was intentionally selected to make model fitting by the package more challenging.

#### **Source**

Claudia Seiler, TU Dresden, Institute of Hydrobiology.

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extcoef_logistic Extended Set of Coefficients of a Logistic Growth Model	
--	--

### **Description**

Estimate model-specific derived parameters of the logistic growth model

# Usage

```
extcoef_logistic(object, quantile = 0.95, time = NULL, ...)
```

# **Arguments**

object model object fited by fit\_growthmodel

quantile fraction of the capacity parameter (K) for the quantile method

time 2-valued vector of the search interval for the independent variable (time). Note:

this needs to be set this manually if saturation is not reached within the observa-

tion time period taken from the data.

... reserved for future extensions

#### **Details**

This function returns the estimated parameters of a logistic growth model (y0, mumax, K) and a series of estimates for the time of approximate saturation. The estimates are defined as follows:

- turnpoint: time of turnpoint (50% saturation)
- sat1: time of the minimum of the 2nd derivative
- sat2: time of the intercept between the steepest increase (the tangent at mumax) and the carrying capacity K
- sat3: time when a quantile of K (default 0.95) is reached

This function is normally not directly called by the user. It is usually called indirectly from coef or results if extended=TRUE.

#### Value

vector that contains the fitted parameters and some derived characteristics (extended parameters) of the logistic function.

#### Note

The estimates for the turnpoint and the time of approximate saturation (sat1, sat2, sat3) may be unreliable, if saturation is not reached within the observation time period. See example below. A set of extended parameters exists currently only for the standard logistic growth model (grow\_logistic). The code and naming of the parameters is preliminary and may change in future versions.

extcoef\_logistic 13

```
## The 'extended parameters' are usually derived
data(antibiotic)
## fit a logistic model to a single data set
dat <- subset(antibiotic, conc==0.078 & repl=="R4")</pre>
parms <- c(y0=0.01, mumax=0.2, K=0.5)
fit <- fit_growthmodel(grow_logistic, parms, dat$time, dat$value)</pre>
coef(fit, extended=TRUE)
## fit the logistic to all data sets
myData <- subset(antibiotic, repl=="R3")</pre>
parms <- c(y0=0.01, mumax=0.2, K=0.5)
all <- all_growthmodels(value ~ time | conc,
                        data = myData, FUN=grow_logistic,
                        p = parms, ncores = 2)
par(mfrow=c(3,4))
plot(all)
results(all, extended=TRUE)
## we see that the the last 3 series (10...12) do not go into saturation
## within the observation time period.
## We can try to extend the search range:
results(all[10:12], extended=TRUE, time=c(0, 5000))
## visualisation how the 'extended parameters' are derived
# Derivatives of the logistic:
   The 1st and 2nd derivatives are internal functions of the package.
   They are used here for the visualisation of the algorithm.
deriv1 <- function(time, y0, mumax, K) {</pre>
 ret <- (K*mumax*y0*(K - y0)*exp(mumax * time))/</pre>
    ((K + y0 * (exp(mumax * time) - 1))^2)
 unname(ret)
}
deriv2 <- function(time, y0, mumax, K) {</pre>
 ret <- -(K * mumax^2 * y0 * (K - y0) * exp(mumax * time) *
            (-K + y0 * exp(mumax * time) + y0))/
    (K + y0 * (exp(mumax * time) - 1))^3
 unname(ret)
}
```

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```
data(bactgrowth)
## extract one growth experiment by name
dat <- multisplit(bactgrowth, c("strain", "conc", "replicate"))[["D:0:1"]]</pre>
## unconstraied fitting
p \leftarrow c(y0 = 0.01, mumax = 0.2, K = 0.1) # start parameters
fit1 <- fit_growthmodel(FUN = grow_logistic, p = p, dat$time, dat$value)</pre>
summary(fit1)
p <- coef(fit1, extended=TRUE)</pre>
## copy parameters to separate variables to improve readability -------
y0 <- p["y0"]
mumax <- p["mumax"]</pre>
K <- p["K"]
turnpoint <- p["turnpoint"]</pre>
sat1 <- p["sat1"] # 2nd derivative</pre>
sat2 <- p["sat2"] # intercept between steepest increase and K
sat3 <- p["sat3"] # a given quantile of K, default 95\%</pre>
## show saturation values in growth curve and 1st and 2nd derivatives -----
opar <- par(no.readonly=TRUE)</pre>
par(mfrow=c(3, 1), mar=c(4,4,0.2,0))
plot(fit1)
## 95% saturation
abline(h=0.95*K, col="magenta", lty="dashed")
## Intercept between steepest increase and 100% saturation
b <- deriv1(turnpoint, y0, mumax, K)</pre>
a <- K/2 - b*turnpoint
abline(a=a, b=b, col="orange", lty="dashed")
abline(h=K, col="orange", lty="dashed")
points(sat2, K, pch=16, col="orange")
points(turnpoint, K/2, pch=16, col="blue")
## sat2 is the minimum of the 2nd derivative
abline(v=c(turnpoint, sat1, sat2, sat3),
       col=c("blue", "grey", "orange", "magenta"), lty="dashed")
## plot the derivatives
with(dat, plot(time, deriv1(time, y0, mumax, K), type="l", ylab="y'"))
abline(v=c(turnpoint, sat1), col=c("blue", "grey"), lty="dashed")
with(dat, plot(time, deriv2(time, y0, mumax, K), type="1", ylab="y''"))
abline(v=sat1, col="grey", lty="dashed")
par(opar)
```

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fit\_easylinear

Fit Exponential Growth Model with a Heuristic Linear Method

### **Description**

Determine maximum growth rates from the log-linear part of a growth curve using a heuristic approach similar to the "growth rates made easy"-method of Hall et al. (2013).

#### Usage

```
fit_easylinear(time, y, h = 5, quota = 0.95)
```

#### **Arguments**

time vector of independent variable.

y vector of dependent variable (concentration of organisms).

h width of the window (number of data).

quota part of window fits considered for the overall linear fit (relative to max. growth

rate)

### **Details**

The algorithm works as follows:

- 1. Fit linear regressions to all subsets of h consecutive data points. If for example h = 5, fit a linear regression to points  $1 \dots 5, 2 \dots 6, 3 \dots 7$  and so on. The method seeks the highest rate of exponential growth, so the dependent variable is of course log-transformed.
- 2. Find the subset with the highest slope  $b_{max}$  and include also the data points of adjacent subsets that have a slope of at least  $quota \cdot b_{max}$ , e.g. all data sets that have at least 95% of the maximum slope.
- 3. Fit a new linear model to the extended data window identified in step 2.

### Value

object with parameters of the fit. The lag time is currently estimated as the intersection between the fit and the horizontal line with  $y=y_0$ , where y0 is the first value of the dependent variable. The intersection of the fit with the abscissa is indicated as y0\_lm (lm for linear model). These identifieres and their assumptions may change in future versions.

### References

```
Hall, BG., Acar, H, Nandipati, A and Barlow, M (2014) Growth Rates Made Easy. Mol. Biol. Evol. 31: 232-38, doi:10.1093/molbev/mst187
```

#### See Also

```
Other fitting functions: all_easylinear(), all_growthmodels(), all_splines(), fit_growthmodel(), fit_spline()
```

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### **Examples**

```
data(bactgrowth)

splitted.data <- multisplit(bactgrowth, c("strain", "conc", "replicate"))
dat <- splitted.data[[1]]

plot(value ~ time, data=dat)
fit <- fit_easylinear(dat$time, dat$value)

plot(fit)
plot(fit, log="y")
plot(fit, which="diagnostics")

fitx <- fit_easylinear(dat$time, dat$value, h=8, quota=0.95)

plot(fit, log="y")
lines(fitx, pch="+", col="blue")

plot(fit)
lines(fitx, pch="+", col="blue")</pre>
```

fit\_growthmodel

Fit Nonlinear Parametric Growth Model

# **Description**

Determine maximum growth rates by fitting nonlinear models.

# Usage

```
fit_growthmodel(
  FUN,
  p,
  time,
  y,
  lower = -Inf,
  upper = Inf,
  which = names(p),
  method = "Marq",
  transform = c("none", "log"),
  control = NULL,
  ...
)
```

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### **Arguments**

FUN	function of growth model to be fitted.
р	named vector of start parameters and initial values of the growth model.
time	vector of independent variable.
у	vector of dependent variable (concentration of organisms).
lower	lower bound of the parameter vector (optional).
upper	upper bound of the parameter vector (optional).
which	vector of parameter names that are to be fitted.
method	character vector specifying the optimization algorithm (see modFit).
transform	fit model to non-transformed or log-transformed data.
control	A list of control parameters for the optimizers. See Details.
	additional parameters passed to the optimizer.

#### **Details**

This function calls modFit from package FME. Syntax of control parameters and available options may differ, depending on the optimizer used, except control=list(trace=...) that switches tracing on and off for all methods and is either TRUE, or FALSE, or an integer value like 0, 1, 2, 3, depending on the optimizer.

#### Value

object with parameters of the fit.

#### See Also

```
modFit about constrained fitting of models to data
Other fitting functions: all_easylinear(), all_growthmodels(), all_splines(), fit_easylinear(), fit_spline()
```

```
data(bactgrowth)
splitted.data <- multisplit(bactgrowth, c("strain", "conc", "replicate"))
## get one element either by index or by name
dat <- splitted.data[[1]]
dat <- splitted.data[["D:0:1"]]

p <- c(y0 = 0.01, mumax = 0.2, K = 0.1)

## unconstraied fitting
fit1 <- fit_growthmodel(FUN = grow_logistic, p = p, dat$time, dat$value)
coef(fit1)
summary(fit1)

## optional box-constraints</pre>
```

fit\_spline

```
lower <- c(y0 = 1e-6, mumax = 0, K = 0)
upper <- c(y0 = 0.05, mumax = 5, K = 0.5)
fit1 <- fit_growthmodel(
   FUN = grow_logistic, p = p, dat$time, dat$value,
   lower = lower, upper = upper)
plot(fit1, log="y")</pre>
```

fit\_spline

Fit Exponential Growth Model with Smoothing Spline

# Description

Determine maximum growth rates from the first derivative of a smoothing spline.

### Usage

```
fit_spline(time, y, optgrid = length(time), ...)
```

# Arguments

time vector of independent variable.

y vector of dependent variable (concentration of organisms).

optgrid number of steps on the x-axis used for the optimum search . algorithm. The

default should work in most cases, as long as the data are equally spaced. A smaller number may lead to non-detectable speed-up, but has the risk that the

search gets trapped in a local minimum.

... other parameters passed to smooth.spline, see details.

### **Details**

The method was inspired by an algorithm of Kahm et al. (2010), with different settings and assumptions. In the moment, spline fitting is always done with log-transformed data, assuming exponential growth at the time point of the maximum of the first derivative of the spline fit.

All the hard work is done by function smooth.spline from package **stats**, that is highly user configurable. Normally, smoothness is automatically determined via cross-validation. This works well in many cases, whereas manual adjustment is required otherwise, e.g. by setting spar to a fixed value [0,1] that also disables cross-validation.

### Value

object with parameters of the fit

#### References

Kahm, M., Hasenbrink, G., Lichtenberg-Frate, H., Ludwig, J., Kschischo, M. 2010. grofit: Fitting Biological Growth Curves with R. Journal of Statistical Software, 33(7), 1-21, doi:10.18637/jss.v033.i07

### See Also

```
Other fitting functions: all_easylinear(), all_growthmodels(), all_splines(), fit_easylinear(), fit_growthmodel()
```

# Examples

```
data(bactgrowth)
splitted.data <- multisplit(bactgrowth, c("strain", "conc", "replicate"))</pre>
dat <- splitted.data[[2]]</pre>
time <- dat$time
     <- dat$value
## automatic smoothing with cv
res <- fit_spline(time, y)</pre>
plot(res, log="y")
plot(res)
coef(res)
## a more difficult data set
dat <- splitted.data[[56]]</pre>
time <- dat$time
y <- dat$value
## default parameters
res <- fit_spline(time, y)</pre>
plot(res, log="y")
## small optgrid, trapped in local minimum
res <- fit_spline(time, y, optgrid=5)</pre>
plot(res, log="y")
## manually selected smoothing parameter
res <- fit_spline(time, y, spar=.5)</pre>
plot(res, log="y")
plot(res, ylim=c(0.005, 0.03))
```

function\_growthmodel-class

Union Class of Growth Model or Function

### **Description**

This class union comprises parametric model functions from class growthmodel and ordinary functions to describe time-dependent growth of organisms.

20 growthmodel

### See Also

the constructor function growthmodel how to create instances of class growthmodel.

growthmodel

Create a User-defined Parametric Growth Model

# **Description**

This constructor method allows to create user-defined functions that can be used as parametric models describing time-dependent growth of organisms.

### Usage

```
growthmodel(x, pnames = NULL)
```

# **Arguments**

 $\boldsymbol{x}$  a function with arguments times and parms, and returning a matrix with two

columns time and y.

pnames character vector with the names of the model parameters.

### **Details**

Package **growthrates** has a plug-in architecture allowing user-defined growth models of the following form:

```
identifier <- function(time, parms) {
    ... content of function here ...
    return(as.matrix(data.frame(time=time, y=y)))
}</pre>
```

where time is a numeric vector and parms a named, non-nested list of model parameters. The constructor function growthmodel is used to attach the names of the parameters as an optional attribute.

#### See Also

```
Other growth models: grow_baranyi(), grow_exponential(), grow_gompertz(), grow_gompertz2(), grow_huang(), grow_logistic(), grow_richards(), ode_genlogistic(), ode_twostep()
```

growthmodel-class 21

### **Examples**

```
test <- function(time, parms) {
  with(as.list(parms), {
    y <- (K * y0) / (y0 + (K - y0) * exp(-mumax * time)) + y_shift
    return(as.matrix(data.frame(time=time, y=y)))
  })
}

mygrowthmodel <- growthmodel(test, c("y0", "mumax", "K", "y_shift"))</pre>
```

growthmodel-class

Class of Growth Model Functions

# **Description**

This class is used for the parametric grow\_... functions of the package and can also be used for user-defined functions to describe time-dependent growth of organisms.

### See Also

the constructor function growthmodel how to create instances of this class.

```
growthrates_fit-class S4 Classes of Package growthrates
```

### **Description**

growthrates\_fit: top-level class representing a growthrates fit with any method.

### **Slots**

```
FUN model function used.

fit results of the model fit.

obs observation data used for model fitting.

rsquared coefficient of determination.

par parameters of the fit.

ndx index values of the time points used (for easylinear_fit).

xy x and y values at the maximum of the spline.
```

22 grow\_baranyi

grow\_baranyi

The Baranyi and Roberts Growth Model

### **Description**

The growth model of Baranyi and Roberts (1995) written as analytical solution of the system of differential equations.

### Usage

```
grow_baranyi(time, parms)
```

### **Arguments**

time

vector of time steps (independent variable).

parms

named parameter vector of the Baranyi growth model with:

- y0 initial value of abundance,
- mumax maximum growth rate (1/time),
- K carrying capacity (max. abundance),
- h0 parameter specifying the initial physiological state of organisms (e.g. cells) and in consequence the lag phase (h0 = max growth rate \* lag phase).

### **Details**

The version of the equation used in this package has the following form:

```
A = time + 1/mumax * log(exp(-mumax * time) + exp(-h0) - exp(-mumax * time - h0))
log(y) = log(y0) + mumax * A - log(1 + (exp(mumax * A) - 1)/exp(log(K) - log(y0)))
```

#### Value

vector of dependent variable (y).

#### References

Baranyi, J. and Roberts, T. A. (1994). A dynamic approach to predicting bacterial growth in food. International Journal of Food Microbiology, 23, 277-294.

Baranyi, J. and Roberts, T.A. (1995). Mathematics of predictive microbiology. International Journal of Food Microbiology, 26, 199-218.

# See Also

```
Other growth models: grow_exponential(), grow_gompertz(), grow_gompertz2(), grow_huang(), grow_logistic(), grow_richards(), growthmodel, ode_genlogistic(), ode_twostep()
```

grow\_exponential 23

### **Examples**

```
time <- seq(0, 30, length=200)
y <- grow_baranyi(time, c(y0=0.01, mumax=.5, K=0.1, h0=5))[,"y"]
plot(time, y, type="1")
plot(time, y, type="1", log="y")</pre>
```

grow\_exponential

Exponential Growth Model

### **Description**

Unlimited exponential growth model.

### Usage

```
grow_exponential(time, parms)
```

### **Arguments**

time

vector of time steps (independent variable).

parms

named parameter vector of the exponential growth model with:

- y0 initial abundance (e.g. concentration of bacterial cells).
- mumax maximum growth rate (1/time).

### **Details**

The equation used is:

```
y = y0 * exp(mumax * time)
```

#### Value

vector of dependent variable (y).

#### See Also

```
Other growth models: grow_baranyi(), grow_gompertz(), grow_gompertz2(), grow_huang(), grow_logistic(), grow_richards(), growthmodel, ode_genlogistic(), ode_twostep()
```

```
time <- seq(0, 30, length=200)
y <- grow_exponential(time, c(y0=1, mumax=0.5))[,"y"]
plot(time, y, type="1")
```

24 grow\_gompertz

grow\_gompertz

Growth Model According to Gompertz

### **Description**

Gompertz growth model written as analytical solution of the differential equation system.

### Usage

```
grow_gompertz(time, parms)
```

### **Arguments**

time vector of time steps (independent variable).

parms named parameter vector of the Gompertz growth model with:

• y0 initial value of abundance,

• mumax maximum growth rate (1/time),

• K maximum abundance (carrying capacity).

#### **Details**

The equation used here is:

```
y = K * exp(log(y0/K) * exp(-mumax * time))
```

#### Value

vector of dependent variable (y)

#### Note

The naming of parameter "mumax" was done in analogy to the other growth models, but it turned out that it was not consistent with the maximum growth rate of the population. This can be considered as bug. The function will be removed or replaced in future versions of the package. Please use grow\_gompertz2 instead.

#### References

Tsoularis, A. (2001) Analysis of Logistic Growth Models. Res. Lett. Inf. Math. Sci, (2001) 2, 23-46.

# See Also

```
Other growth models: grow_baranyi(), grow_exponential(), grow_gompertz2(), grow_huang(), grow_logistic(), grow_richards(), growthmodel, ode_genlogistic(), ode_twostep()
```

grow\_gompertz2 25

### **Examples**

grow\_gompertz2

Growth Model According to Gompertz

### **Description**

Gompertz growth model written as analytical solution of the differential equation system.

# Usage

```
grow_gompertz2(time, parms)
grow_gompertz3(time, parms)
```

# **Arguments**

time

vector of time steps (independent variable).

parms

named parameter vector of the Gompertz growth model with:

- y0 initial value of abundance,
- mumax maximum growth rate (1/time),
- K maximum abundance (carrying capacity),
- lambda time of lag phase of the 3 parameter Gompertz model .

### **Details**

The equation used here is:

```
y = y0*(K/y0)^(exp(-exp((exp(1)*mumax*(lambda-time))/log(K/y0)+1)))
```

Functions grow\_gompert2 and grow\_gompertz3 describe sigmoidal growth with an exponentially decreasing intrinsic growth rate with or without an additional lag parameter. The formula follows the reparametrization of Zwietering et al (1990), with parameters that have a biological meaning.

#### Value

vector of dependent variable (y)

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### References

Tsoularis, A. (2001) Analysis of Logistic Growth Models. Res. Lett. Inf. Math. Sci, (2001) 2, 23-46.

Zwietering, M. H., Jongenburger, I., Rombouts, F. M., and Van't Riet, K. (1990). Modeling of the bacterial growth curve. Appl. Environ. Microbiol., 56(6), 1875-1881.

#### See Also

```
Other growth models: grow_baranyi(), grow_exponential(), grow_gompertz(), grow_huang(), grow_logistic(), grow_richards(), growthmodel, ode_genlogistic(), ode_twostep()
```

### **Examples**

```
time <- seq(0, 30, length=200)
y <- grow_gompertz(time, c(y0=1, mumax=.2, K=10))[,"y"]
plot(time, y, type="1", ylim=c(0, 12))</pre>
```

grow\_huang

Growth Model According to Huang

# **Description**

Huangs growth model written as analytical solution of the differential equations.

### Usage

```
grow_huang(time, parms)
```

### **Arguments**

time

vector of time steps (independent variable).

parms

named parameter vector of Huang's growth model with:

- y0 initial value of abundance,
- mumax maximum growth rate (1/time),
- K carrying capacity (max. total concentration of cells),
- alpha shape parameter determining the curvature,
- lambda parameter determining the lag time.

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### **Details**

The version of the equation used in this package has the following form:

```
B = time + 1/alpha * log((1 + exp(-alpha * (time - lambda)))/(1 + exp(alpha * lambda))) log(y) = log(y0) + log(K) - log(y0 + (K - y0) * exp(-mumax * B))
```

In contrast to the original publication, all parameters related to population abundance (y, y0, K) are given as untransformed values. They are not log-transformed.

In general, using log-transformed parameters would indeed be a good idea to avoid the need of constained optimization, but tests showed that box-constrained optimization worked resonably well. Therefore, handling of optionally log-transformed parameters was removed from the package to avoid confusion. If you want to discuss this, please let me know.

#### Value

vector of dependent variable (y).

#### References

Huang, Lihan (2008) Growth kinetics of Listeria monocytogenes in broth and beef frankfurters - determination of lag phase duration and exponential growth rate under isothermal conditions. Journal of Food Science 73(5), E235 – E242. doi:10.1111/j.17503841.2008.00785.x

Huang, Lihan (2011) A new mechanistic growth model for simultaneous determination of lag phase duration and exponential growth rate and a new Belehdradek-type model for evaluating the effect of temperature on growth rate. Food Microbiology 28, 770 – 776. doi:10.1016/j.fm.2010.05.019

Huang, Lihan (2013) Introduction to USDA Integrated Pathogen Modeling Program (IPMP). Residue Chemistry and Predictive Microbiology Research Unit. USDA Agricultural Research Service.

### See Also

```
Other growth models: grow_baranyi(), grow_exponential(), grow_gompertz(), grow_gompertz2(), grow_logistic(), grow_richards(), growthmodel, ode_genlogistic(), ode_twostep()
```

```
time <- seq(0, 30, length=200)
y <- grow_huang(time, c(y0=0.01, mumax=.1, K=0.1, alpha=1.5, lambda=3))[,"y"]
plot(time, y, type="1")
plot(time, y, type="1", log="y")</pre>
```

28 grow\_logistic

grow\_logistic

Logistic Growth Model

### **Description**

Classical logistic growth model written as analytical solution of the differential equation.

### Usage

```
grow_logistic(time, parms)
```

# **Arguments**

time

vector of time steps (independent variable)

parms

named parameter vector of the logistic growth model with:

- yo initial value of population measure
- mumax intrinsic growth rate (1/time)
- K carrying capacity (max. total concentration of cells)

### **Details**

The equation used is:

$$y = (K * y0)/(y0 + (K - y0) * exp(-mumax * time))$$

#### Value

vector of dependent variable (y).

### See Also

```
Other growth models: grow_baranyi(), grow_exponential(), grow_gompertz(), grow_gompertz2(), grow_huang(), grow_richards(), growthmodel, ode_genlogistic(), ode_twostep()
```

```
time <- seq(0, 30, length=200)
y <- grow_logistic(time, c(y0=1, mumax=0.5, K=10))[,"y"]
plot(time, y, type="1")
```

grow\_richards 29

grow\_richards

Growth Model According to Richards

### **Description**

Richards growth model written as analytical solution of the differential equation.

### Usage

```
grow_richards(time, parms)
```

#### **Arguments**

time

vector of time steps (independent variable).

parms

named parameter vector of the Richards growth model with:

- y0 initial value of abundance,
- mumax maximum growth rate (note different interpretation compared to exponential growth),
- K carrying capacity (max. total concentration of cells),
- beta shape parameter determining the curvature.

### **Details**

The equation used is:

$$y = K * (1 - exp(-beta * mumax * time) * (1 - (y0/K)^{-}beta))^{(-1/beta)}$$

The naming of parameters used here follows the convention of Tsoularis (2001), but uses mumax for growthrate and y for abundance to make them consistent to other growth functions.

### Value

vector of dependent variable (y).

### References

Richards, F. J. (1959) A Flexible Growth Function for Empirical Use. Journal of Experimental Botany 10 (2): 290–300.

Tsoularis, A. (2001) Analysis of Logistic Growth Models. Res. Lett. Inf. Math. Sci, (2001) 2, 23–46.

### See Also

```
Other growth models: grow_baranyi(), grow_exponential(), grow_gompertz(), grow_gompertz2(), grow_huang(), grow_logistic(), growthmodel, ode_genlogistic(), ode_twostep()
```

names.growthmodel

### **Examples**

lm\_or\_NULL-class

Union Class of Linear Model or NULL

### **Description**

Class to handle no-growth cases

names.growthmodel

Get Names Attributes of Growth Models

### **Description**

Methods to get the parameter names of a growth model or to get or set identifiers of multiple\_fits objects.

### Usage

```
## $3 method for class 'growthmodel'
names(x)
## $4 method for signature 'multiple_fits'
names(x)
## $4 replacement method for signature 'multiple_fits'
names(x) <- value</pre>
```

### **Arguments**

x either a function being a parametric growth model of package **growthmodels** or

an object with multiple fits.

value a character vector of up to the same length as x, or NULL

### Value

character vector of the parameter names

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### Methods

**Method for class** growthmodel: returns information about valid parameter names if a pnames attribute exists, else NULL. NULL.

**Method for class** multiple\_fits: can be applied to objects returned by all\_growthmodels, all\_splines or all\_easylinear respectively. This can be useful for selecting subsets, e.g. for plotting, see example below.

### See Also

```
multiple_fits, all_growthmodels, all_splines, all_easylinear
```

### **Examples**

ode\_genlogistic

Generalized Logistic Growth Model

# Description

Generalized logistic growth model solved as differential equation.

### Usage

```
ode_genlogistic(time, y, parms, ...)
grow_genlogistic(time, parms, ...)
```

# Arguments

time vector of simulation time steps
y named vector with initial value of the system (e.g. cell concentration)
parms parameters of the generalized logistic growth model
• mumax maximum growth rate (1/time)

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- K carrying capacity (max. abundance)
- alpha, beta, gamma parameters determining the shape of growth. Setting all values to one returns the ordinary logistic function.

additional parameters passed to the ode-function.

#### **Details**

The model is given as its first derivative:

$$dy/dt = mumax * y^a lpha * (1 - (y/K)^b eta)^g amma$$

that is then numerically integrated ('simulated') according to time (t).

The generalized logistic according to Tsoularis (2001) is a flexible model that covers exponential and logistic growth, Richards, Gompertz, von Bertalanffy, and some more as special cases.

The differential equation is solved numerically, where function ode\_genlogistic is the differential equation, and grow\_genlogistic runs a numerical simulation over time.

The default version grow\_genlogistic is run directly as compiled code, whereas the R versions ode\_logistic is provided for testing by the user.

#### Value

For ode\_genlogistic: matrix containing the simulation outputs. The return value of has also class deSolve.

For grow\_genlogistic: vector of dependent variable (y).

- time time of the simulation
- y abundance of organisms

#### References

Tsoularis, A. (2001) Analysis of Logistic Growth Models. Res. Lett. Inf. Math. Sci, (2001) 2, 23-46.

#### See Also

```
Other growth models: grow_baranyi(), grow_exponential(), grow_gompertz(), grow_gompertz2(), grow_huang(), grow_logistic(), grow_richards(), growthmodel, ode_twostep()
```

```
time <- seq(0, 30, length=200)
parms <- c(mumax=0.5, K=10, alpha=1, beta=1, gamma=1)
y0 <- c(y=.1)
out <- ode(y0, time, ode_genlogistic, parms)
plot(out)

out2 <- ode(y0, time, ode_genlogistic, parms = c(mumax=0.2, K=10, alpha=2, beta=1, gamma=1))
out3 <- ode(y0, time, ode_genlogistic, parms = c(mumax=0.2, K=10, alpha=1, beta=2, gamma=1))</pre>
```

ode\_twostep 33

```
out4 <- ode(y0, time, ode_genlogistic, parms = c(mumax=0.2, K=10, alpha=1, beta=1, gamma=2))
out5 <- ode(y0, time, ode_genlogistic, parms = c(mumax=0.2, K=10, alpha=.5, beta=1, gamma=1))
out6 <- ode(y0, time, ode_genlogistic, parms = c(mumax=0.2, K=10, alpha=1, beta=.5, gamma=1))
out7 <- ode(y0, time, ode_genlogistic, parms = c(mumax=0.3, K=10, alpha=1, beta=1, gamma=.5))
plot(out, out2, out3, out4, out5, out6, out7)

## growth with lag (cf. log_y)
plot(ode(y0, time, ode_genlogistic, parms = c(mumax=1, K=10, alpha=2, beta=.8, gamma=5)))</pre>
```

ode\_twostep

Twostep Growth Model

# **Description**

System of two differential equations describing bacterial growth as two-step process of activation (or adaptation) and growth.

### Usage

```
ode_twostep(time, y, parms, ...)
grow_twostep(time, parms, ...)
```

### **Arguments**

time actual time (for the ode) resp. vector of simulation time steps.

y named vector with state of the system (yi, ya: abundance of inactive and active

organisms, e.g. concentration of inactive resp. active cells).

parms parameters of the two-step growth model:

- yi, ya initial abundance of active and inactive organisms,
- kw activation ("wakeup") constant (1/time),
- mumax maximum growth rate (1/time),
- K carrying capacity (max. abundance).

placeholder for additional parameters (for user-extended versions of this function)

### **Details**

The model is given as a system of two differential equations:

$$dy_i/dt = -kw * yi$$

$$dy_a/dt = kw * yi + mumax * (1 - (yi + ya)/K) * ya$$

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that are then numerically integrated ('simulated') according to time (t). The model assumes that the population consists of active  $(y_a)$  and inactive  $(y_i)$  cells so that the observed abundance is  $(y = y_i + y_a)$ . Adapting inactive cells change to the active state with a first order 'wakeup' rate (kw).

Function ode\_twostep is the system of differential equations, whereas grow\_twostep runs a numerical simulation over time.

A similar two-compartment model, but without the logistic term, was discussed by Baranyi (1998).

#### Value

For ode\_twostep: matrix containing the simulation outputs. The return value of has also class deSolve.

For grow\_twostep: vector of dependent variable (y):

- time time of the simulation
- yi concentration of inactive cells
- ya concentration of active cells
- y total cell concentration

#### References

Baranyi, J. (1998). Comparison of stochastic and deterministic concepts of bacterial lag. J. heor. Biol. 192, 403–408.

#### See Also

```
Other growth models: grow_baranyi(), grow_exponential(), grow_gompertz(), grow_gompertz2(), grow_huang(), grow_logistic(), grow_richards(), growthmodel, ode_genlogistic()

Other growth models: grow_baranyi(), grow_exponential(), grow_gompertz(), grow_gompertz2(), grow_huang(), grow_logistic(), grow_richards(), growthmodel, ode_genlogistic()
```

plot 35

|--|

# Description

Methods to plot growth model fits together with the data and, alternatively, plot diagnostics

#### **Usage**

```
## S4 method for signature 'nonlinear_fit,missing'
plot(x, y, log = "", which = c("fit", "diagnostics"), ...)

## S4 method for signature 'nonlinear_fit'
lines(x, ...)

## S4 method for signature 'easylinear_fit,missing'
plot(x, y, log = "", which = c("fit", "diagnostics"), ...)

## S4 method for signature 'smooth.spline_fit,missing'
plot(x, y, ...)

## S4 method for signature 'easylinear_fit'
lines(x, ...)

## S4 method for signature 'multiple_fits,missing'
plot(x, y, ...)
```

# Arguments

X	an object returned by a model fitting function of package <b>growthrates</b> , that can contain one or multiple fits.
У	(ignored) for compatibility with the default plot method.
log	a character string which contains "y" if the y axis is to be logarithmic.
which	either "fit" (default) or "diagnostics".
	other arguments pased to the plotting methods, see plot default and par.

### **Details**

The plot methods detect automatically which type of plot is appropriate, depending on the class of x and can plot either one single model fit or a complete series (multiple fits). In the latter case it may be wise to redirect the graphics to an external file (e.g. a pdf) and / or to use tomething like par(mfrow=c(3,3)).

The lines-method is currently only available for single fits.

If you need more control, you can of course also write own plotting functions.

### See Also

```
plot.default, par, fit_growthmodel, fit_easylinear, all_growthmodels, all_easylinear
```

# **Examples**

```
data(bactgrowth)
splitted.data <- multisplit(bactgrowth, c("strain", "conc", "replicate"))</pre>
## get table from single experiment
dat <- splitted.data[["D:0:1"]]</pre>
fit1 <- fit_spline(dat$time, dat$value)</pre>
plot(fit1, log="y")
plot(fit1)
## derive start parameters from spline fit
p <- coef(fit1)</pre>
## subset of first 10 data
first10 <- dat[1:10, ]
fit2 <- fit_growthmodel(grow_exponential, p=p, time=first10$time, y=first10$value)</pre>
p <- c(coef(fit1), K = max(dat$value))</pre>
fit3 <- fit_growthmodel(grow_logistic, p=p, time=dat$time, y=dat$value, transform="log")</pre>
plot(fit1)
lines(fit2, col="green")
lines(fit3, col="red")
all.fits <- all_splines(value ~ time | strain + conc + replicate, data = bactgrowth)
par(mfrow=c(3,3))
plot(all.fits)
## it is also possible to plot a single fit or a subset of the fits
par(mfrow=c(1,1))
plot(all.fits[["D:0:1"]])
par(mfrow=c(2,2))
plot(all.fits[1:4])
## plot only the 'R' strain
par(mfrow=c(4, 6))
plot(all.fits[grep("R:", names(all.fits))])
```

predict,growthrates\_fit-method

Model Predictions for growthrates Fits

### **Description**

Class-specific methods of package **growthrates** to make predictions.

### Usage

```
## S4 method for signature 'growthrates_fit'
predict(object, ...)
## S4 method for signature 'smooth.spline_fit'
predict(object, newdata = NULL, ..., type = c("exponential", "spline"))
## S4 method for signature 'easylinear_fit'
predict(object, newdata = NULL, ..., type = c("exponential", "no_lag"))
## S4 method for signature 'nonlinear_fit'
predict(object, newdata, ...)
## S4 method for signature 'multiple_fits'
predict(object, ...)
```

# Arguments

object name of a 'growthrates' object for which prediction is desired. additional arguments affecting the predictions produced. an optional data frame with column 'time' for new time steps with which to newdata predict. type of predict. Can be 'exponential' or 'spline' for fit\_spline, resp. type

'exponential' or 'no\_lag' for fit\_easylinear.

### **Details**

The implementation of the predict methods is still experimental and under discussion.

#### See Also

```
methods, predict.smooth.spline, predict.lm, predict.nls
```

```
data(bactgrowth)
splitted.data <- multisplit(bactgrowth, c("strain", "conc", "replicate"))</pre>
## get table from single experiment
dat <- splitted.data[[1]]</pre>
## --- linear fit ------
fit <- fit_easylinear(dat$time, dat$value)</pre>
plot(fit)
```

```
pr <- predict(fit)</pre>
lines(pr[,1:2], col="blue", lwd=2, lty="dashed")
pr <- predict(fit, newdata=list(time=seq(2, 6, .1)), type="no_lag")</pre>
lines(pr[,1:2], col="magenta")
## --- spline fit -------------
fit1 <- fit_spline(dat$time, dat$value, spar=0.5)</pre>
coef(fit1)
summary(fit1)
plot(fit1)
pr <- predict(fit1)</pre>
lines(pr[,1:2], lwd=2, col="blue", lty="dashed")
pr <- predict(fit1, newdata=list(time=2:10), type="spline")</pre>
lines(pr[,1:2], lwd=2, col="cyan")
## --- nonlinear fit ------
dat <- splitted.data[["T:0:2"]]</pre>
p <- c(y0 = 0.02, mumax = .5, K = 0.05, h0 = 1)
fit2 <- fit_growthmodel(grow_baranyi, p=p, time=dat$time, y=dat$value)</pre>
## prediction for given data
predict(fit2)
## prediction for new data
pr <- predict(fit2, newdata=data.frame(time=seq(0, 50, 0.1)))</pre>
plot(fit2, xlim=c(0, 50))
lines(pr[, c("time", "y")], lty="dashed", col="red")
```

rsquared,growthrates\_fit-method

Accessor Methods of Package growthrates.

### **Description**

Functions to access the results of fitted growthrate objects: summary, coef, rsquared, deviance, residuals, df.residual, obs, results.

# Usage

```
## S4 method for signature 'growthrates_fit'
rsquared(object, ...)
## S4 method for signature 'growthrates_fit'
obs(object, ...)
```

```
## S4 method for signature 'growthrates_fit'
coef(object, extended = FALSE, ...)
## S4 method for signature 'easylinear_fit'
coef(object, ...)
## S4 method for signature 'smooth.spline_fit'
coef(object, extended = FALSE, ...)
## S4 method for signature 'growthrates_fit'
deviance(object, ...)
## S4 method for signature 'growthrates_fit'
summary(object, ...)
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summary(object, cov = TRUE, ...)
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residuals(object, ...)
## S4 method for signature 'growthrates_fit'
df.residual(object, ...)
## S4 method for signature 'smooth.spline_fit'
summary(object, cov = TRUE, ...)
## S4 method for signature 'smooth.spline_fit'
df.residual(object, ...)
## S4 method for signature 'smooth.spline_fit'
deviance(object, ...)
## S4 method for signature 'multiple_fits'
coef(object, ...)
## S4 method for signature 'multiple_fits'
rsquared(object, ...)
## S4 method for signature 'multiple_fits'
deviance(object, ...)
## S4 method for signature 'multiple_fits'
results(object, ...)
## S4 method for signature 'multiple_easylinear_fits'
results(object, ...)
```

```
## S4 method for signature 'multiple_fits'
summary(object, ...)
## S4 method for signature 'multiple_fits'
residuals(object, ...)
```

### **Arguments**

object name of a 'growthrate' object.
... other arguments passed to the methods.

extended boolean if extended set of parameters shoild be printed cov boolean if the covariance matrix should be printed.

### **Examples**

```
data(bactgrowth)
splitted.data <- multisplit(bactgrowth, c("strain", "conc", "replicate"))

## get table from single experiment
dat <- splitted.data[[10]]

fit1 <- fit_spline(dat$time, dat$value, spar=0.5)
coef(fit1)
summary(fit1)

## derive start parameters from spline fit
p <- c(coef(fit1), K = max(dat$value))
fit2 <- fit_growthmodel(grow_logistic, p=p, time=dat$time, y=dat$value, transform="log")
coef(fit2)
rsquared(fit2)
deviance(fit2)
summary(fit2)

plot(residuals(fit2) ~ obs(fit2)[,2])</pre>
```

# **Description**

Operators to access parts of 'multiple\_fits' objects

# Usage

```
## S4 method for signature 'multiple_fits,ANY,missing'
x[i, j, ..., drop = TRUE]
## S4 method for signature 'multiple_fits,ANY,missing'
x[[i, j, ...]]
```

# Arguments

x	object of class multiple_fits
i	numeric or character index
j	NULL (for compatibility with other uses of [ or [[)
	optional arguments passed to [
drop	If TRUE the result is coerced to the lowest possible dimension

```
data(bactgrowth)
L <- all_splines(value ~ time | strain + conc +replicate, data=bactgrowth)

coef(L[[1]])
plot(L[["R:0:2"]])

par(mfrow=c(2, 2))
plot(L[1:4])</pre>
```

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